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Regulation of Spire1 by Phosphoinositides and PARP in DNA Damage Response

C 1 0 Undergraduate Research Fellowship Programme (URFP); BIOC4999 Biochemistry Project

Participant: Xiangyu Ouyang (UID: 3035551539) Supervisor: Prof. Michael Huen Keywords: DNA Damage Response; Phosphoinositides; Spire1; PARP; IPMK; Nuclear Actin Polymerization

Abstract: Phosphoinositides (PPIs) plays an essential role in regulating nuclear actin polymerization upon DNA damage. SpireI, an actin nucleator, was reported to be required for actin filament formation and DNA repair. Interestingly, SpireI harbors a putative PPI-binding FYVE domain, which was previously shown to swiftly enrich at DNA lesions. Here, I investigated whether PPIs and PPI-related enzymes regulates SpireI-FYVE accumulation at laser-induced DNA breaks. Upon DNA damage, nuclear PPIs [PtdIns(4)P and PtdIns(4,5)P2] co-localized with SpireI-FYVE, which was repressed by inositol polyphosphate multikinase (IPMK) inhibition. Furthermore, suppressed PARP activity attenuated the accumulation of SpireI-FYVE and PPIs at DNA damage sites. To-

gether, my results establish a potential regulatory arm of nuclear actin via the PARP/IPMK/PPI axis in response to DNA damage.

Introduction

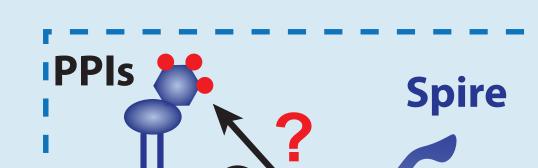
DNA repair via actin and PPIs

Actin polymerization actively participates in various DNA damage response (DDR) processes, including chromatin modulation, DNA break translocation, and repair factor recruitment. Recent findings revealed **1** poly-phosphoinositides (PPIs) enriched at double-strand breaks (DSBs) **2** to stimulate actin filament formation **3** and ATR signaling **4** to facilitate DNA repair. Further, **5** functions of PPIs in DDR are determined by the activity of inositol polyphosphate multikinase (IPMK), a nuclear kinase converting PtdIns(4,5)P2 to PIP3. However, the molecular basis of how PPIs regulate nuclear actin remains largely enigmatic.

Spire1 – a potential link

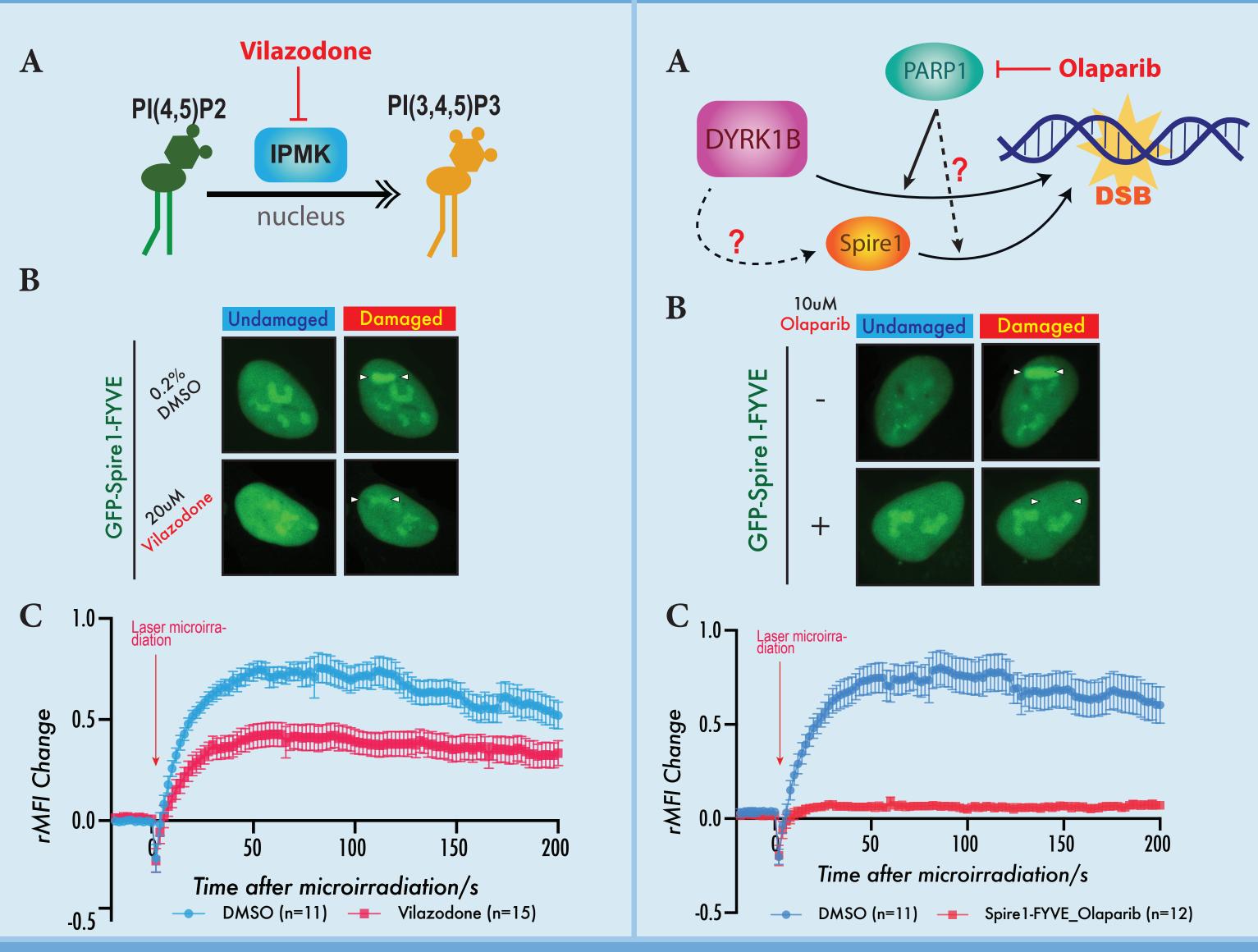
Spire1 belongs to the **Spire actin nucleator** family, which **6** initiates the assembly of monomeric actin into filamentous polymers. Previous studies illustrated Spire1/2 were required for DDR-directed actin polymerization, and Spire1 was also identified as a substrate of DYRK1B, a DDR kinase. Further, our lab demonstrated the accumulation of full-length Spire1 at laser-induced DSBs. Intriguingly, Spire1 harbors an **FYVE domain with putative PPI-binding capability**, **7** indicating a **potential Spire1-PPI interaction**.

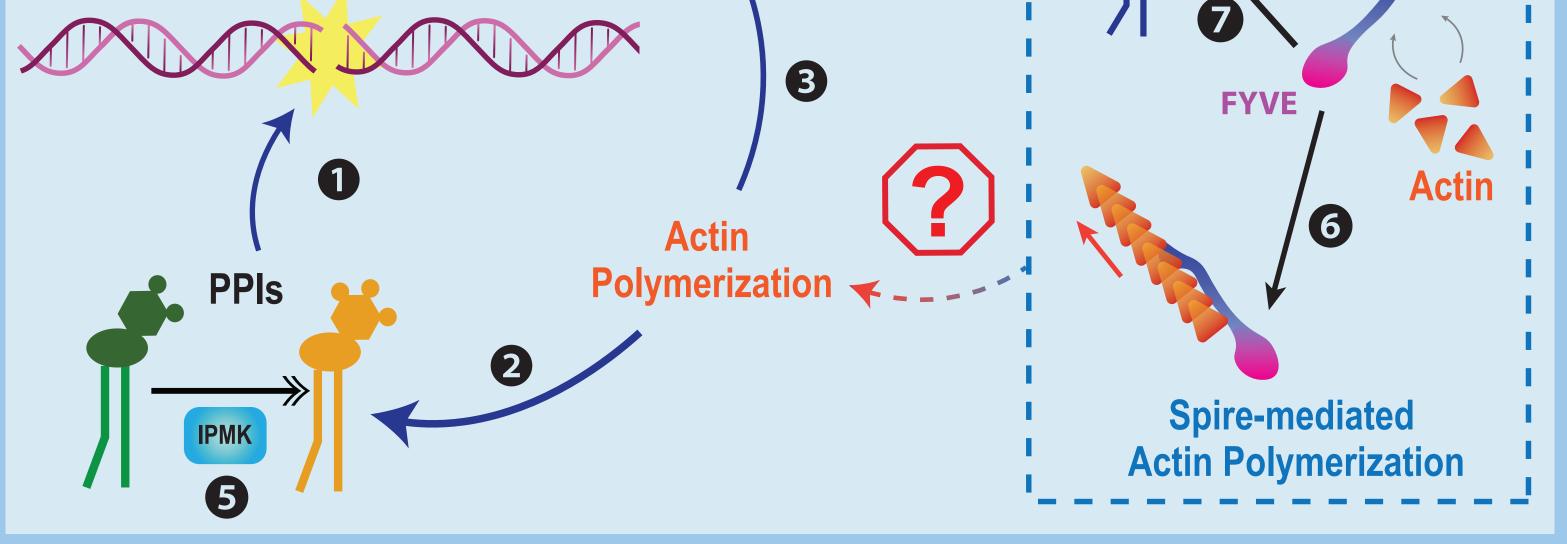




Methods & Results

(2) IPMK inhibition attenuated Spire1-FYVE accumulation at DSB. (3) Inhibiting PARP suppressed recruitments of Spire 1-FYVE.



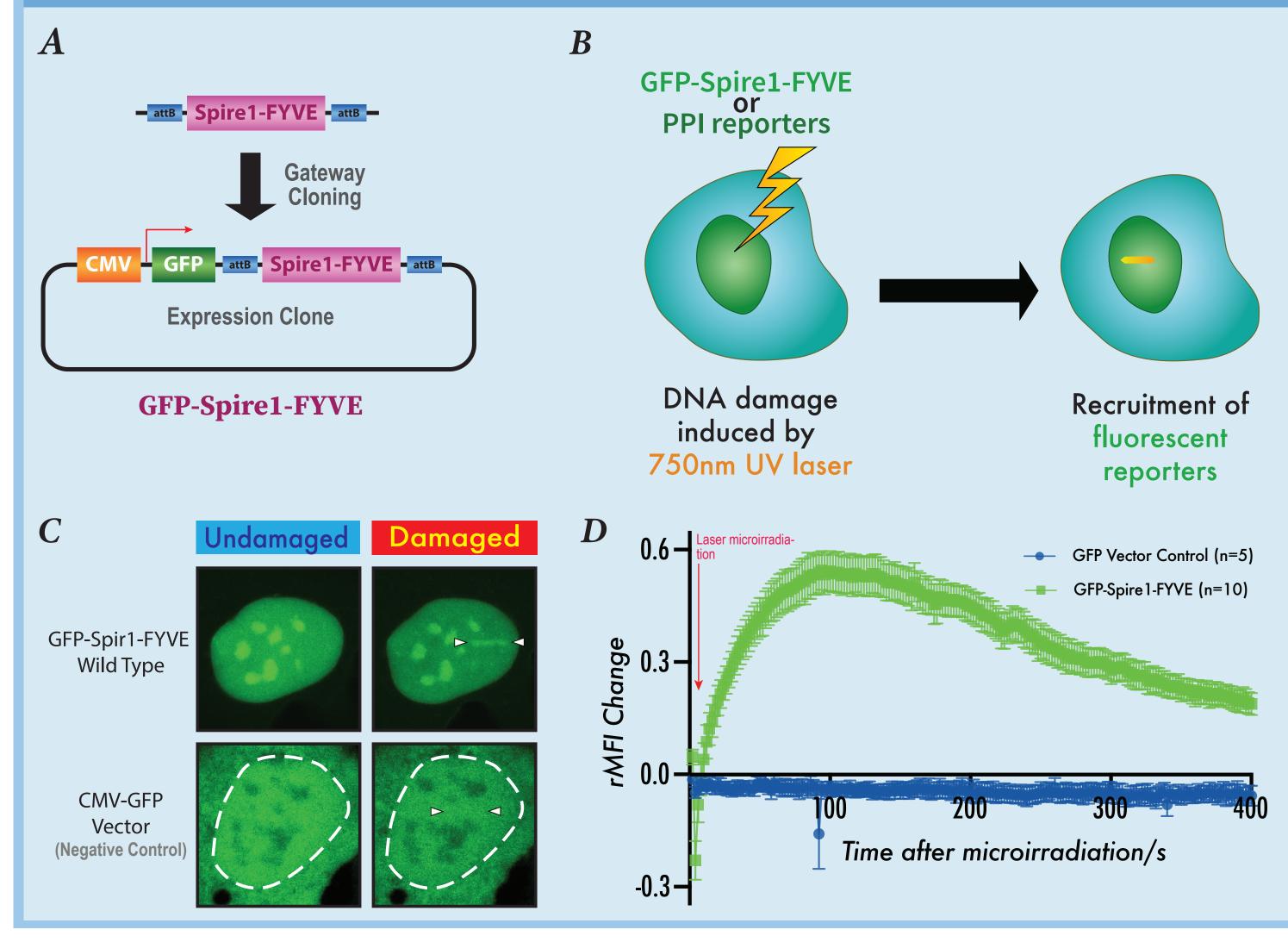


Research Aims

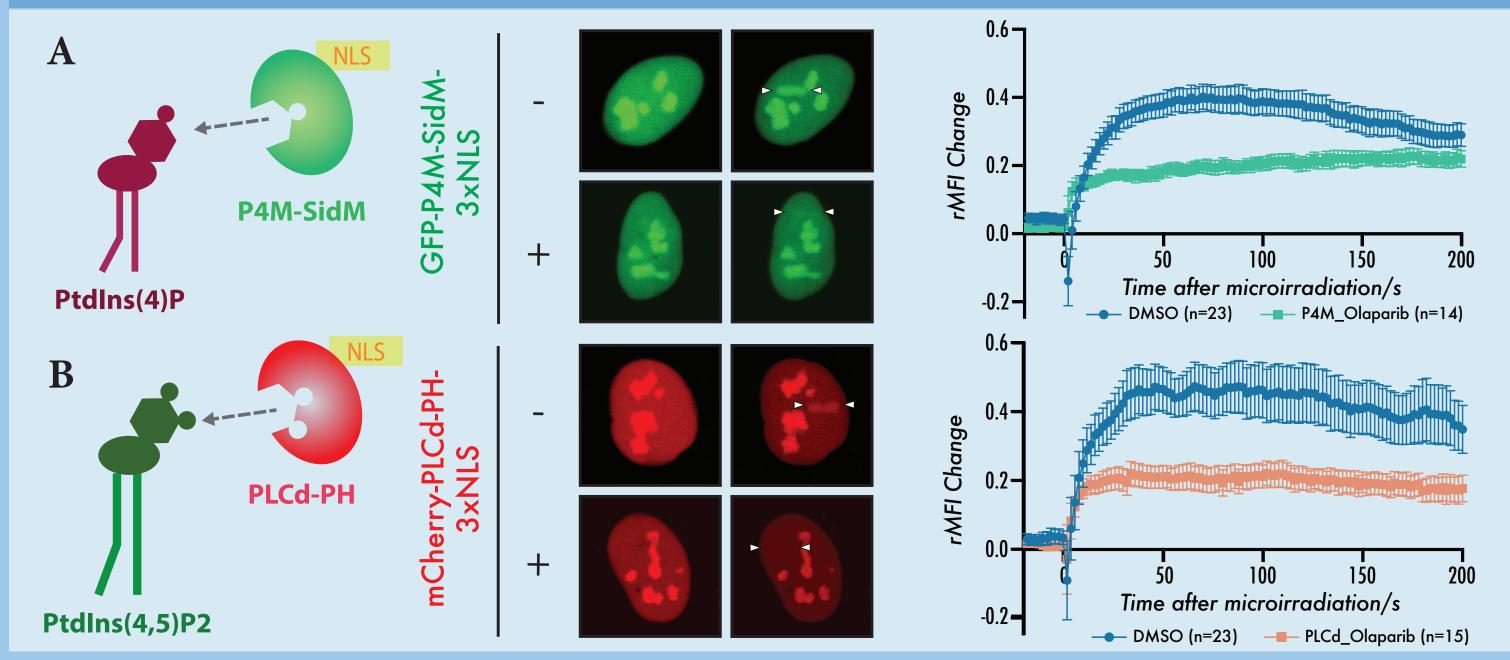
To illustrate DSB-accumulation of Spire1-FYVE
To investigate roles of PPIs and IPMK in the recruitment of Spire1-FYVE
To examine potential upstream regulation of PPIs (e.g., by PARP)

Methods & Results

(1) Accumulation of GFP-Spire1-FYVE at laser-irradiated DNA breaks.



(4) Inhibiting PARP suppressed PPIs enrichment att DSB.



Conclusions

Spire1 recognizes DNA breaks via its FYVE domain.
IPMK is essential for the recruitment of Spire1-FYVE.
Spire1 is likely to interact with PIP3.
Novel role of PARP in regulating actin polymerization via the PPI/Spire axis in DDR.

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References

Baum, B., and Kunda, P. (2005). Actin Nucleation: Spire — Actin Nucleator in a Class of Its Own. Current Biology 15, R305-R308. Belin, B.J., Lee, T., and Mullins, R.D. (2015). DNA damage induces nuclear actin filament assembly by Formin-2 and Spire-1/2 that promotes efficient DNA repair. eLife 4, e07735.

Caridi, C.P., Plessner, M., Grosse, R., and Chiolo, I. (2019). Nuclear actin filaments in DNA repair dynamics. Nature Cell Biology 21, 1068-1077. Dong, C., West Kirk, L., Tan Xin, Y., Li, J., Ishibashi, T., Yu, C.-h., Sy Shirley, M.H., Leung Justin, W.C., and Huen Michael, S.Y. (2020). Screen identifies DYRK1B network as mediator of transcription repression on damaged chromatin. Proceedings of the National Academy of Sciences 117, 17019-17030. Lee, B., Park, S.J., Lee, S., Park, S.E., Lee, E., Song, J.-J., Byun, Y., and Kim, S. (2020). Identification of the Antidepressant Vilazodone as an Inhibitor of Inositol Polyphosphate Multikinase by Structure-Based Drug Repositioning. Mol Cells 43, 222-227.

Thorsell, A.-G., Ekblad, T., Karlberg, T., Löw, M., Pinto, A.F., Trésaugues, L., Moche, M., Cohen, M.S., and Schüler, H. (2017). Structural Basis for Potency and Promiscuity in Poly(ADP-ribose) Polymerase (PARP) and Tankyrase Inhibitors. J Med Chem 60, 1262-1271.

Tittel, J., Welz, T., Czogalla, A., Dietrich, S., Samol-Wolf, A., Schulte, M., Schwille, P., Weidemann, T., and Kerkhoff, E. (2015). Membrane targeting of the Spirformin actin nucleator complex requires a sequential handshake of polar interactions. The Journal of biological chemistry 290, 6428-6444. Wang, Y.-H., Hariharan, A., Bastianello, G., Toyama, Y., Shivashankar, G.V., Foiani, M., and Sheetz, M.P. (2017). DNA damage causes rapid accumulation of phosphoinositides for ATR signaling. Nat Commun 8, 2118.